

# Binding of optically pure (–)-[<sup>3</sup>H]nicotine to rat brain membranes

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With the recent availability of (–)-[<sup>3</sup>H]nicotine of high specific activity, binding studies were performed on rat brain membranes in the presence of a variety of nicotine analogues and cholinergic drugs. Both a higher affinity ( $K_d = 2 \times 10^{-10}$  M) and a lower affinity ( $2 \times 10^{-9}$  M) site were observed; the stereoselectivity of both sites being similar. A good correlation was observed between  $IC_{50}$ -values and psychotropic potency of a series of *N'*-alkyl substituted nicotine analogues.

Nicotine receptor	Rat brain	Nicotine analogue Cholinergic drug	Nicotine psychopharmacology
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## 1. INTRODUCTION

Recently, a number of studies have appeared on the binding of <sup>3</sup>H-labeled nicotine to rat brain membranes [1–5], but with one exception [3] the studies were not performed with the naturally occurring, optically pure (–)-nicotine. With (–)-[<sup>3</sup>H]nicotine having a specific activity of 5 Ci/mmol, authors in [3] obtained (with rat brain membranes) a  $K_d$  of  $5 \times 10^{-6}$  M and a binding density of  $0.01 \times 10^{-14}$  mol/mg protein. With (±)-[<sup>3</sup>H]nicotine of higher specific activity, the  $K_d$ -values ranged from  $6$ – $100 \times 10^{-9}$  M, while stereoselectivity of the (–)-nicotine over (+)-nicotine ranged from 0–20-times [1–5]. With the recent availability of (–)-[<sup>3</sup>H]nicotine of very high specific activity (87 Ci/mmol), we sought to re-examine its binding characteristics to rat brain membranes, particularly with regard to its stereoselectivity.

## 2. MATERIALS AND METHODS

The procedures for preparation of rat brain

membranes and for measuring specific [<sup>3</sup>H]nicotine binding are described in [1]. Binding was determined by a centrifuge assay. To a 2.0 ml polypropylene tube was added 2 mg membrane protein along with various concentrations of (–)-[<sup>3</sup>H]nicotine (spec. act. 87 Ci/mmol; New England Nuclear) with or without various concentrations of (+)- or (–)-nicotine, nicotine analogues, and cholinergic drugs in a final volume of 1.2 ml 0.05 M Tris, pH 7.5. After incubating in an ice bath (0–4°C) for 15 min, the tubes were centrifuged in an Eppendorf centrifuge for 2 min and the pellet washed twice by filling the tubes with Tris (pH 7.5) and aspirating. The bottom of the tubes was then cut off (animal nail clipper) and counted by liquid scintillation.

The optical purity of the (–)-[<sup>3</sup>H]nicotine was determined by the use of *P. putida* (gift of A.M. Tometsko) as in [6]. Preparation of *N'*-alkyl-substituted nicotine derivatives was done by the method in [7] as modified in [8]. Psychotropic potency was measured by administering the agent to rats via cannulae implanted into the lateral ventricles and determining the minimal dose required to induce prostration involving all 4 limbs [1,5].

## 3. RESULTS

A Scatchard plot of specific (–)-[<sup>3</sup>H]nicotine binding was curvilinear and was resolvable into two slopes; yielding a higher affinity  $K_d$ -value of  $1.9 \times 10^{-10}$  M and a lower one of  $1.7 \times 10^{-9}$  M (fig.1).  $B_m$ -values were  $0.51 \times 10^{-14}$  and  $2.9 \times 10^{-14}$  mol/mg membrane protein, respectively.

A variety of nicotine analogues and cholinergic agents were tested for their ability to compete with (–)-[<sup>3</sup>H]nicotine for binding to rat brain membranes (table 1). The data are presented for binding ( $IC_{50}$ ) at both  $2 \times 10^{-10}$  and  $2 \times 10^{-9}$  M [<sup>3</sup>H]nicotine to distinguish binding at the higher and lower affinity sites, respectively. Data on the psychotropic potency of the agents are also presented; the value designating the relative potency to (–)-nicotine to induce prostration in rats. The  $IC_{50}$ -value for (+)-nicotine was 3-times greater than that for (–)-nicotine at the higher affinity site and 2-times greater at the lower affinity

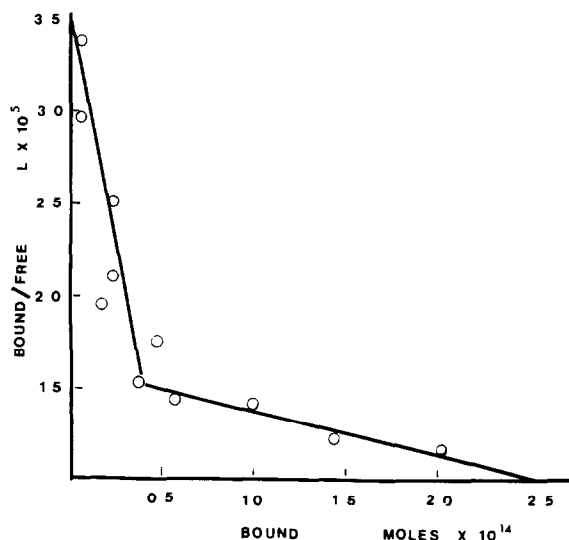


Fig.1. Scatchard plot of specific (–)-[<sup>3</sup>H]nicotine binding.

Table 1

$IC_{50}$ -values against [<sup>3</sup>H]nicotine binding and relative psychopharmacologic potency of various nicotine analogues and cholinergic agents

Agent	High affinity $IC_{50}$ (M)	Lower affinity $IC_{50}$ (M)	Prostration relative potency
(–)-Nicotine	$1 \times 10^{-10}$	$3 \times 10^{-10}$	1
(+)-Nicotine	$3 \times 10^{-10}$	$1 \times 10^{-9}$	1/20
(±)-Nornicotine	$2 \times 10^{-10}$	$1 \times 10^{-9}$	1/20
(±)-N'-Ethylornnicotine	$4 \times 10^{-9}$	$1 \times 10^{-8}$	1/10
(–)-Anabasine	$8 \times 10^{-8}$	$5 \times 10^{-7}$	1/20
(±)-N'-isopropyl nornicotine	$8 \times 10^{-8}$	$6 \times 10^{-7}$	1/40
(±)-N'-n-butyl nornicotine	$2 \times 10^{-7}$	$1 \times 10^{-6}$	IA
(±)-6-β-hydroxyethyl nicotine	$5 \times 10^{-9}$	$3 \times 10^{-8}$	1/30
N-benzyl nornicotine	$1 \times 10^{-8}$	$2 \times 10^{-7}$	IA
Cotinine	M	$> 1 \times 10^{-4}$	IA
Acetylcholine	$1 \times 10^{-6}$	$5 \times 10^{-6}$	IA
Carbamylcholine	$2 \times 10^{-8}$	$8 \times 10^{-7}$	IA
Hexamethonium	$1 \times 10^{-4}$	$1 \times 10^{-4}$	IA
α-Bungarotoxin	$> 5 \times 10^{-5}$	$> 5 \times 10^{-5}$	IA
Mecamylamine	$5 \times 10^{-5}$	$1 \times 10^{-4}$	IA
1-3-Quinuclidinyl benzilate	$8 \times 10^{-5}$	$1 \times 10^{-4}$	IA
Oxotremorine	$5 \times 10^{-5}$	$1 \times 10^{-4}$	IA

$IC_{50}$ -values for higher affinity site were obtained at  $2 \times 10^{-10}$  M [<sup>3</sup>H]nicotine and at  $2 \times 10^{-9}$  M at lower affinity site, and are average of 3 separate experiments agreeing within 7%. Prostration was determined as described in [1,5] and is expressed as relative potency to 1-nicotine which was effective at 30 nmol (in 5 μl). IA = inactive at 50-times the dose of 1-nicotine

site; whereas the psychotropic potency of (–)-nicotine was 20-times that of the (+)-isomer. A 20-fold and 40-fold increase in the  $IC_{50}$ -values occurred with replacement of methyl by ethyl or isopropyl, respectively; and the psychotropic potency was 1/10 and 1/40 that of (–)-nicotine, respectively. Replacement of methyl by *n*-butyl increased the  $IC_{50}$  still further and abolished psychotropic activity. Substitution of  $\beta$ -hydroxyethyl in the 6 position of nicotine increased the  $IC_{50}$  50-fold, while decreasing psychotropic activity 30-fold. Although the binding affinity of nornicotine was 1/2 that of (–)-nicotine, it was only 1/20 as effective as (–)-nicotine in producing prostration. Hexamethonium and  $\alpha$ -bungarotoxin, which are both nicotinic cholinergic antagonists, had  $IC_{50}$ -values around  $1 \times 10^{-4}$  M for both the higher and lower affinity sites. Acetylcholine, as well as the remaining cholinergic agonists and antagonists, also exhibited low binding affinity and did not produce prostration even at 50-times the effective dose of (–)-nicotine.

#### 4. DISCUSSION

The present studies were undertaken to determine whether the binding characteristics of (–)-[ $^3$ H]nicotine to rat brain membranes were significantly different from those obtained with (±)-[ $^3$ H]nicotine. With (±)-[ $^3$ H]nicotine, the affinity of (–)-isomer was 3-times greater than that of (+)-nicotine; however, no greater stereoselectivity was found in the present study using (–)-[ $^3$ H]nicotine. The major new finding was the demonstration of a higher affinity site with a  $K_m$  of  $1.9 \times 10^{-10}$  M and a  $B_m$  of  $0.65 \times 10^{-14}$  mol/mg membrane protein; while the lower affinity site had a  $K_m$  of  $1.9 \times 10^{-9}$  and a  $B_m$  of  $3.0 \times 10^{-14}$  mol/mg, which agreed with previous data with the racemic [ $^3$ H]nicotine [1,5]. Included in this study were some nicotine analogues with modifications in the pyrrolidine *N*. As the alkyl chain length increased from methyl to butyl, there was a dramatic decrease in the binding affinity with a corresponding decrease in psychotropic potency. Replacement of methyl by benzyl decreased binding affinity by over two orders of magnitude; and, although the analogue did not

produce prostration, it acted as an antagonist to the nicotine-induced prostration [1,5]. As reported in [1,2,4], mecamylamine, hexamethonium,  $\alpha$ -bungarotoxin, and other nicotinic cholinergic antagonists exhibited very low binding affinity for the nicotine site and did not produce prostration, with the exception of carbamylcholine and acetylcholine, which had  $IC_{50}$ -values in the  $\mu$ M range. The remainder of cholinergic agents showed very little binding, while none produced prostration at 50-times the dose of nicotine.

Our main conclusions are that the affinity of the (–)-isomer of nicotine to rat brain membranes is no more than 3-times greater than that of the (+)-isomer; the relative binding of the two enantiomers being similar whether (–)-[ $^3$ H]nicotine or (±)-[ $^3$ H]nicotine was used as the ligand. An additional finding was the existence of a higher affinity site; however, the stereoselectivity of this site was no different from that of the lower affinity site. Finally, with a series of *N*'-substituted nicotine derivatives, a good correlation was observed between binding affinity and potency in producing prostration in rats following administration of (–)-nicotine intraventricularly.

#### ACKNOWLEDGEMENT

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#### REFERENCES

- [1] Abood, L.G., Reynolds, D.T. and Bidlack, J.M. (1980) *Life Sci.* 27, 1307–1314.
- [2] Romano, C. and Goldstein, A. (1980) *Science* 210, 647–649.
- [3] Vincek, W.D., Martin, B.R., Aceto, M.D. and Bownan, E.R. (1980) *J. Med. Chem.* 23, 960–962.
- [4] Sershen, H., Reith, M.E.A., Lajtha, A. and Gennara, J. jr (1981) *J. Receptor Res.* 2, 1–15.
- [5] Abood, L.G., Reynolds, D.T., Booth, H. and Bidlack, J.M. (1981) *Neurosci. Biobehav. Rev.* 5, 479–486.
- [6] DeTraglia, M.C. and Tometsko, A. (1980) *Appl. Environ. Microbiol.* 39, 1067–1072.
- [7] Smissman, E.E. and Ruenitz, P.C. (1976) *J. Org. Chem.* 41, 1593–1597.
- [8] Seeman, J.I., Secor, H.V. and Forrest, G. (1979) *J. Labelled Compd. Radiopharm.* 16, 387–395.